

#### **EDMI** Microsystems and Microelectronics

MICRO-614: Electrochemical Nano-Bio-Sensing and Bio/CMOS interfaces

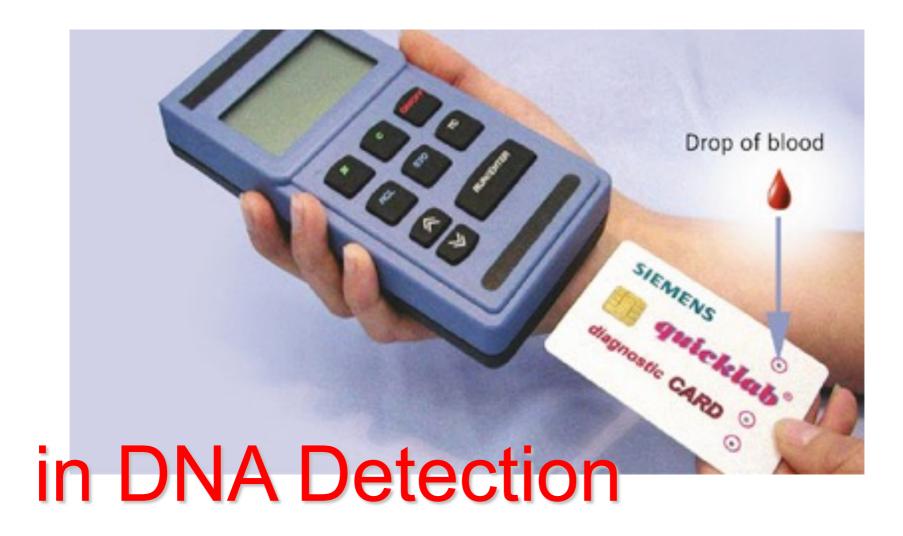
# Lecture #13 CMOS interfaces for DNA Detection

#### **Lecture Outline**

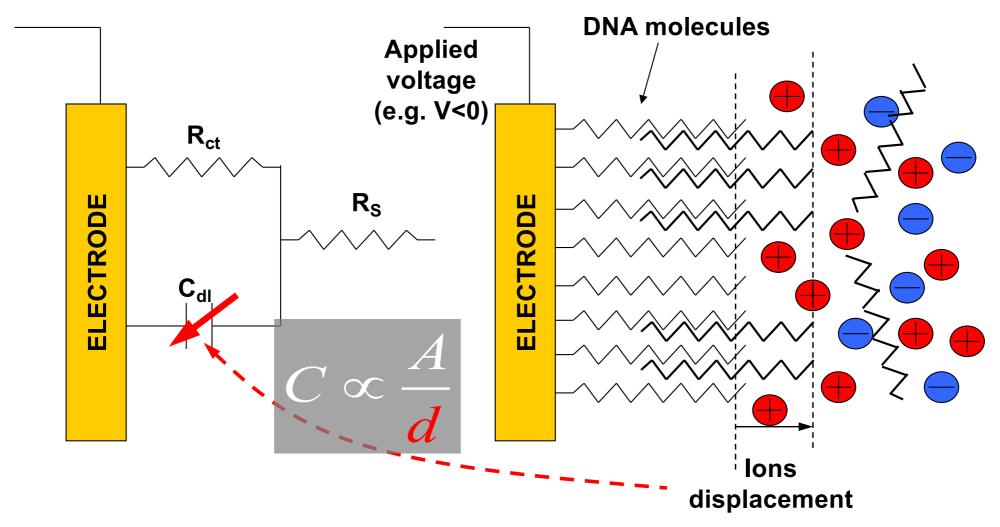
(Book Bio/CMOS: Chapter' paragraphs § 7.1-8)

- CMOS for capacitance detection
- Charge-Based Capacitance Measurement (CBCM)
   Method
- Frequency-to-Capacitance Measurement (FTCM)
   Method

#### **CMOS** architectures for VLSI

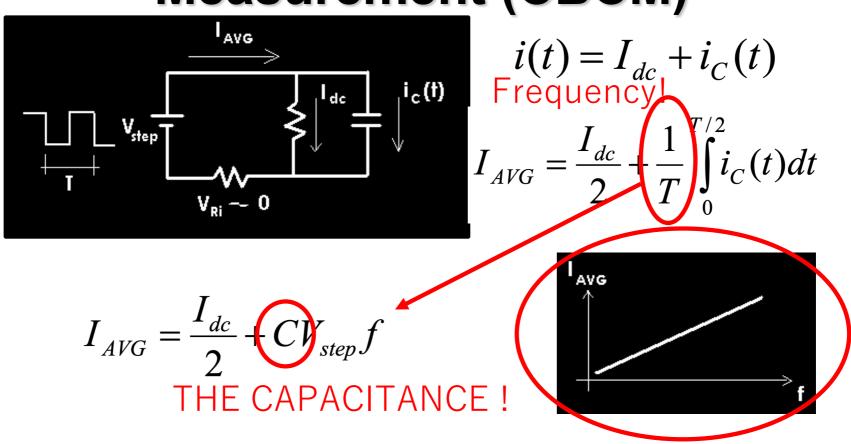


### The Capacitance DNA Detection



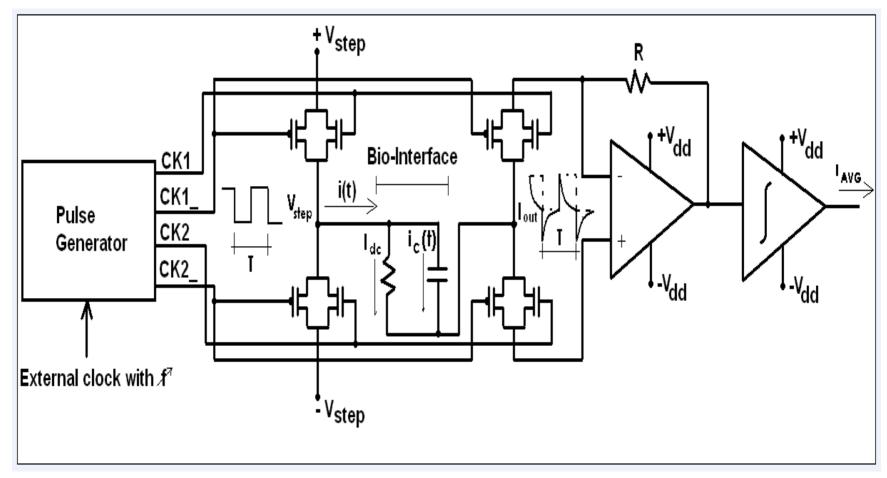
Unlabeled ssDNA may be detected with capacitance measurements as due to charge displacement

# Current Based Capacitance Measurement (CBCM)



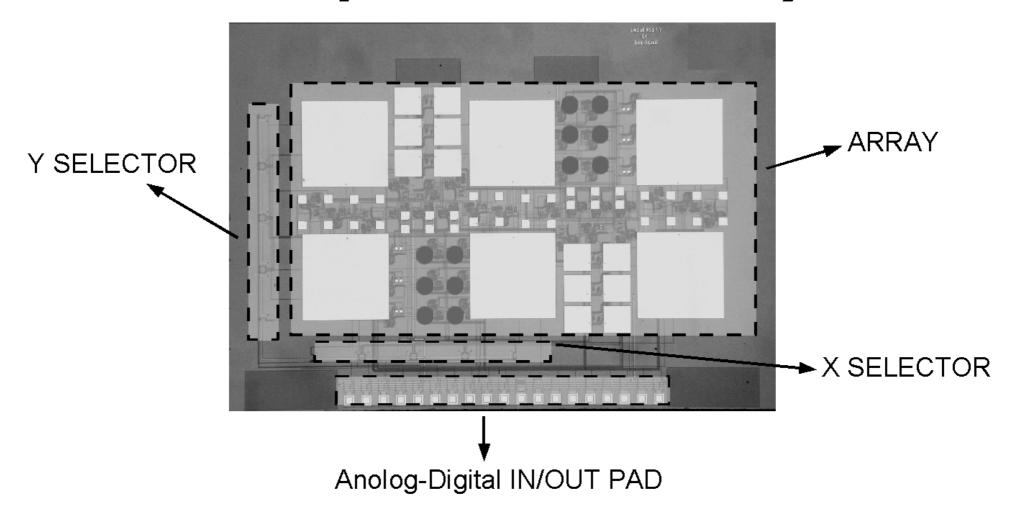
Method for a precise Capacitance measurement

### **CMOS for CBCM detection**



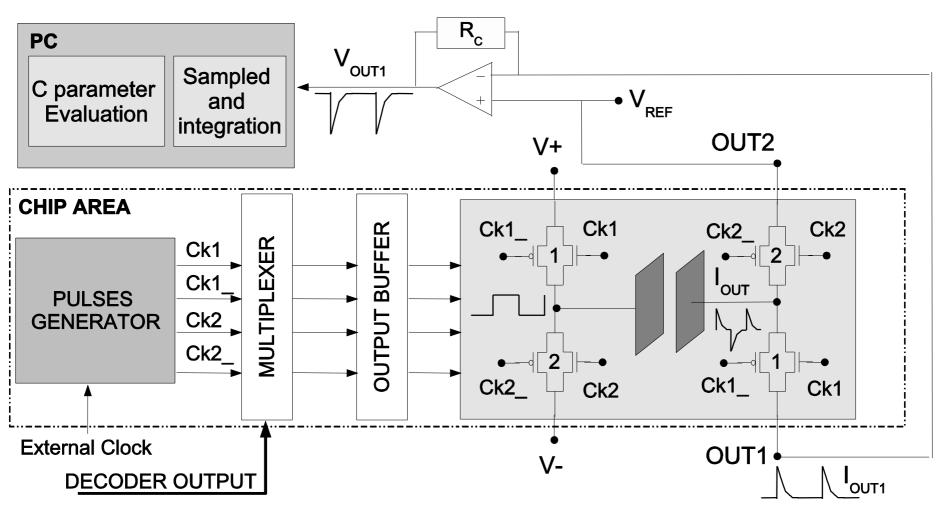
The circuit assures the square signal generator, an inverters, and an integrator to calculate the average current

### The Chip Electrodes Layout



### The VLSI Implementation of the Chip (CBCM method)

(CBCM = Charge Base Capacitance Mode)



### The problem of overlapping signals

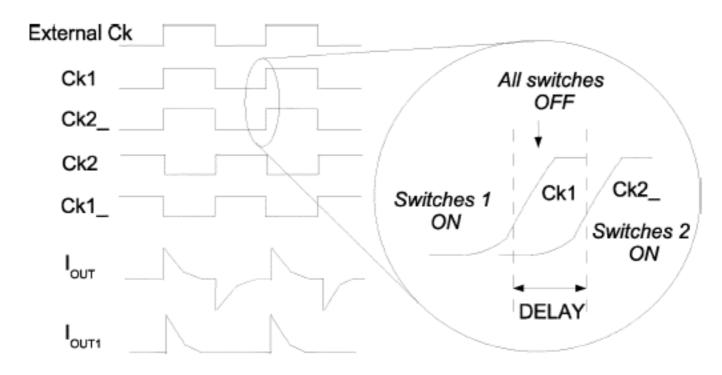


Fig. 4. Schematic representation of the signals flow used in the experiments.

Ck and Ck\_ signals need to be not-overlapping in order to assure the correct square signal generation

### The circuit solution

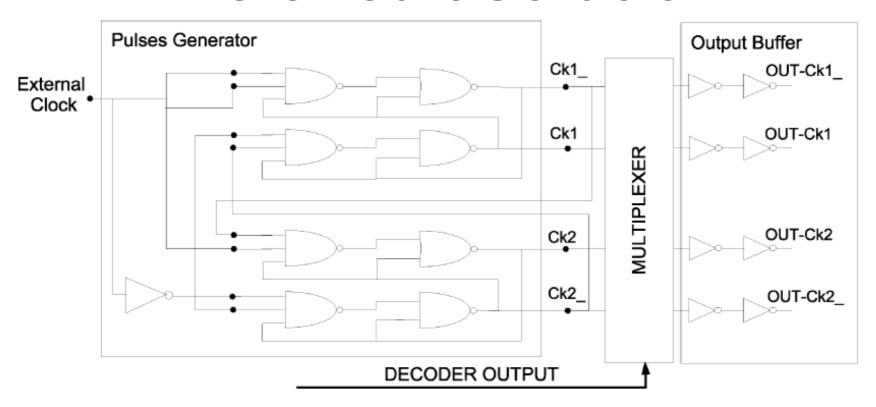


Fig. 5. Schematic plot of the block used to generate not overlapping clock signals.

A simple logical circuit and a digital multiplexer assures not-overlapping Ck and Ck\_ signals

### The Measurements Set-up

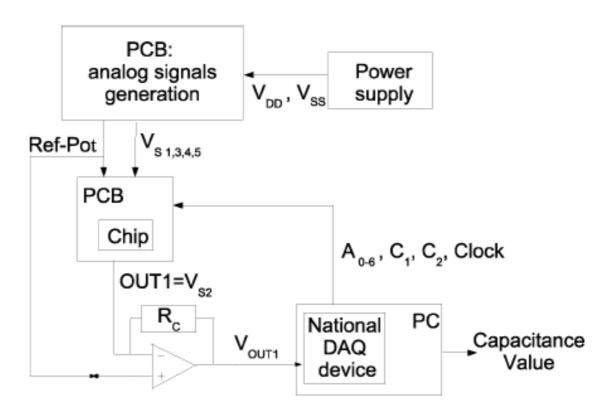
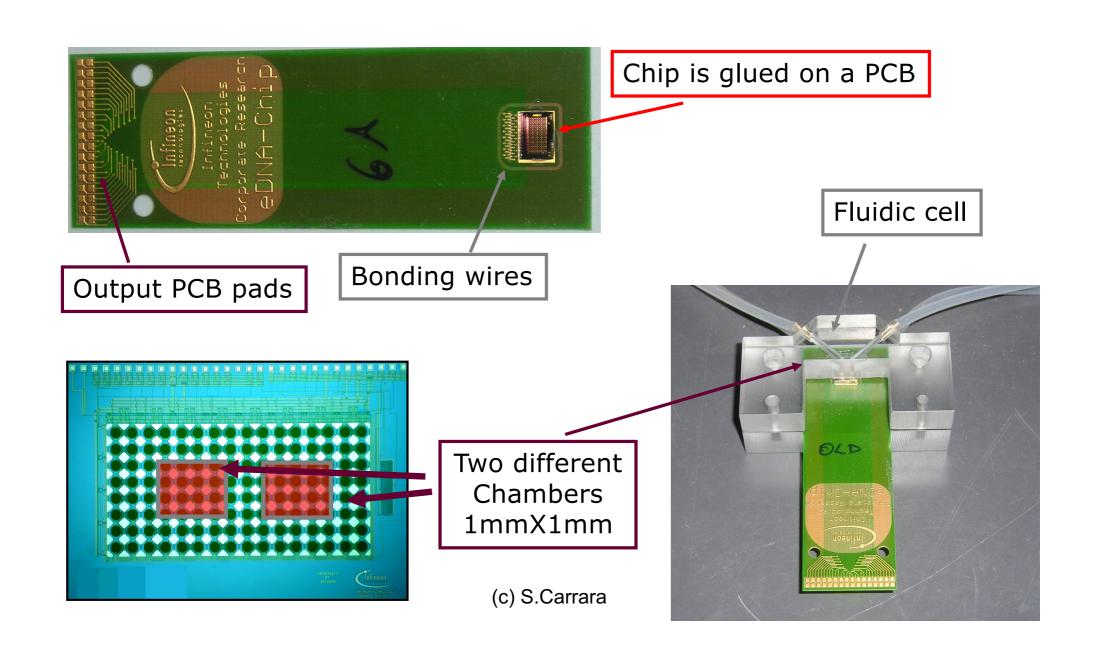


Fig. 8. Schematic representation of the measurement setup.

### The Chip has been mounted onto a PCB for PC remote control and testing

### Liquid Measurement set-up



#### **DNA** detection in CBCM mode

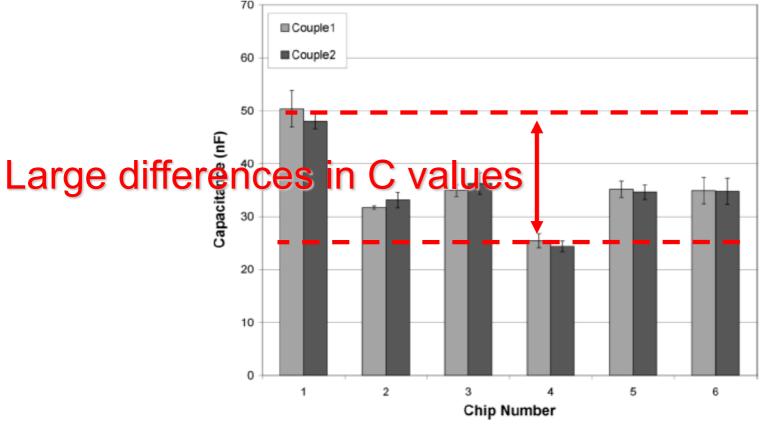


Fig. 10. Capacitance measurements of electrode couples on different chips.

The chip-by-chip reproducibility has been not so high: the problem is on the chip electrodes cleaning

Capacitance vs Frequency

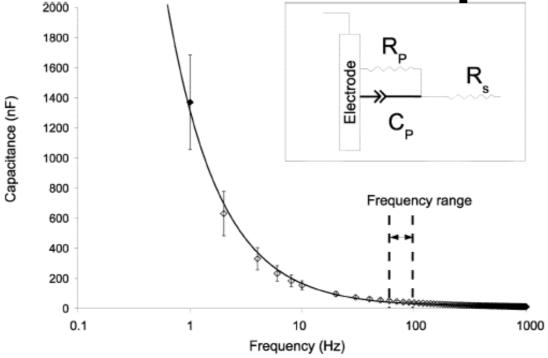
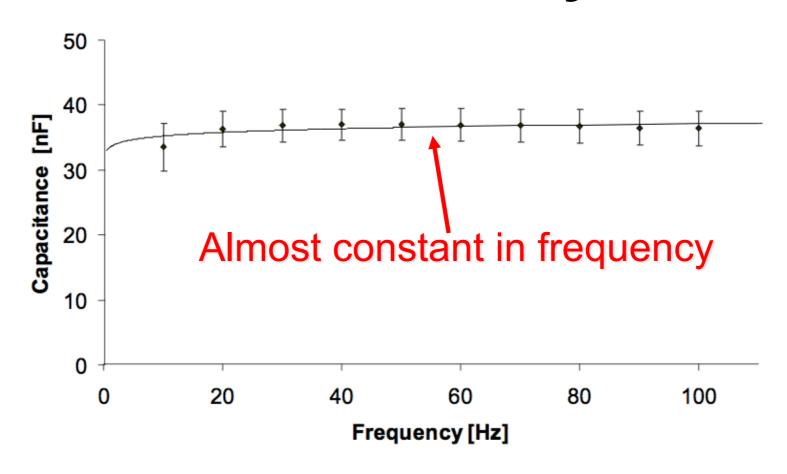


Fig. 9. Measured capacitance versus charge/discharge frequency on clean gold electrodes. The continuous line shows the fitting.

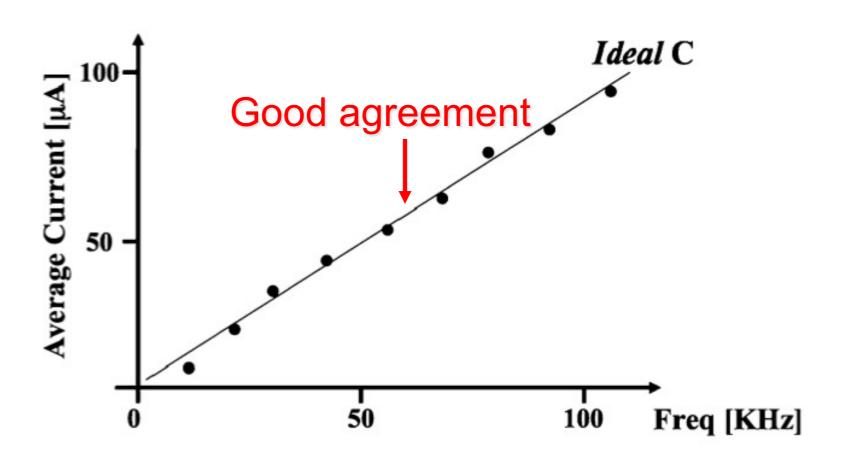
The trends of the measured capacitance vs frequency decrease the accuracy of the measurements in CBCM mode

### **Good DNA Layer**



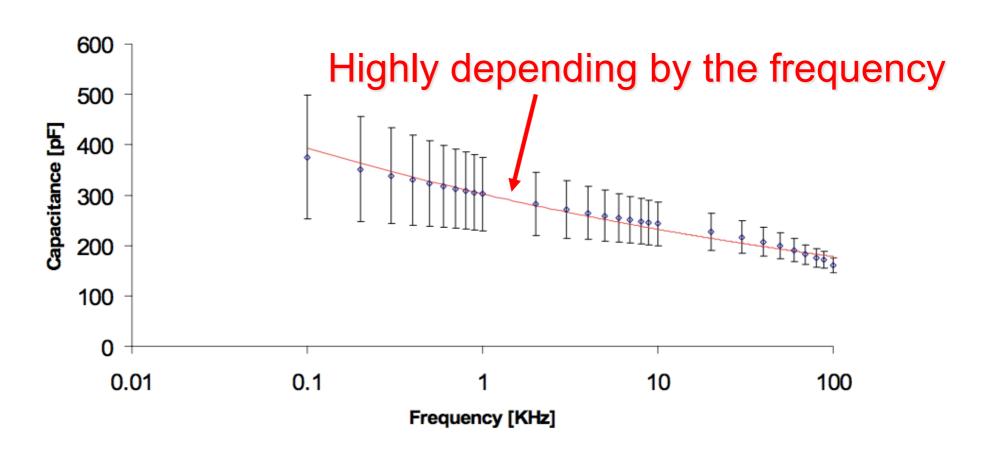
DNA Layer is independent by the frequency thanks to probes immobilized on Ethylene-Glycol Thiols

### **CBCM** on good **DNA** Layer



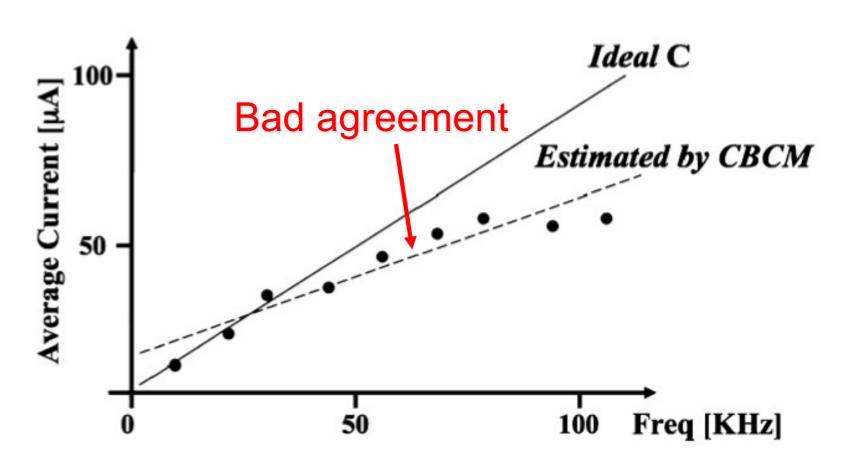
CBCM method on a DNA Layer that is independents by the frequency

### **Bad DNA Layer**



DNA Layer is dependent by the frequency since the monolayer is not extremely well formed

### **CBCM** on bad DNA Layer



CBCM method on a DNA Layer that dependents by the frequency

#### **DNA** detection in CBCM mode

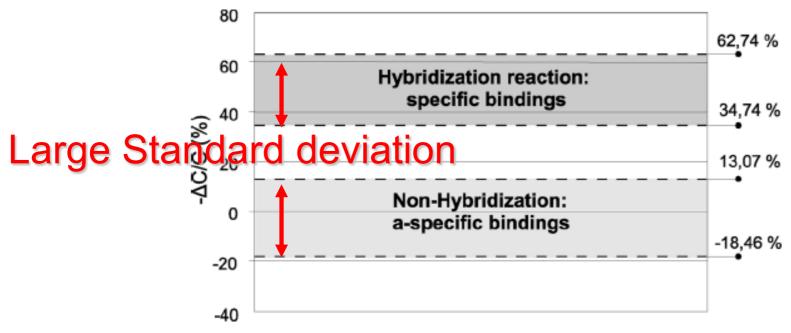


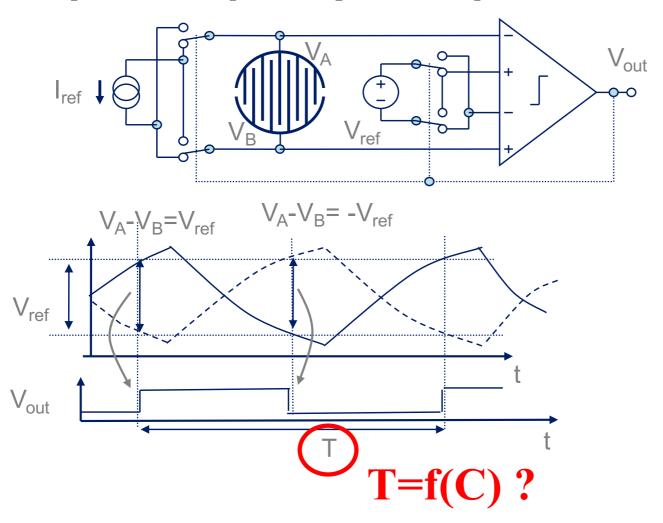
Fig. 12. Capacitance variations due to specific and a-specific bindings (upper and lower bands of measured capacitances, respectively). Positive values indicate capacitance decrease.

The reproducibility on the same chip-spot is not so high: here the problem is on the nano-scale aperture in the probes surfaces

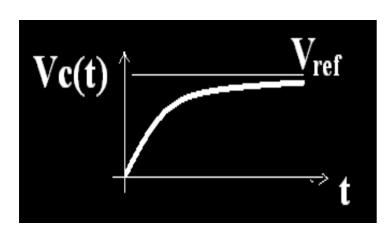
(c) S.Carrara

### Frequency to Capacitance Measurement (FTCM)

Principle: Frequency To Capacitance Mode



## **Current Based Capacitance Measurement (CBCM)**



$$V_c(t) = V_{charge} \left[ 1 - e^{-\frac{t}{RC}} \right]$$

$$V_c(\frac{T}{2}) = V_{ref} = V_{charge} \left[ 1 - e^{-\frac{T}{2RC}} \right]$$

$$V_c(\frac{T}{2}) = V_{ref} = RI_{ref} \left[ 1 - e^{-\frac{T}{2RC}} \right]$$

$$\frac{T}{2RC} = -\ln\left[1 - \frac{V_{ref}}{RI_{ref}}\right] \longrightarrow T = 2RC \ln\left[1 - \frac{V_{ref}}{RI_{ref}}\right]^{-1}$$

### **The Taylor Series**

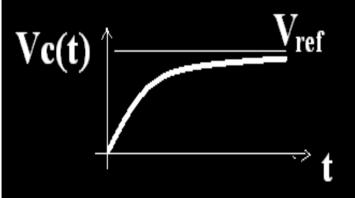
$$f(x) = f(0) + \left\{ \frac{\partial f(x)}{\partial x} \right\} x + \left\{ \frac{\partial^2 f(x)}{\partial x^2} \right\} x^2 + o(3)$$

$$\frac{1}{1-x} = 1 + \{-(-1)\}x + o(2) \cong 1 + x$$

$$\ln\left[\frac{1}{1-x}\right] = \ln[1+x+o(2)] \cong \ln[1+x] = 0+x+o(2) \cong x$$

Linearity by approximation in the right range of values

## Current Based Capacitance Measurement (CBCM)



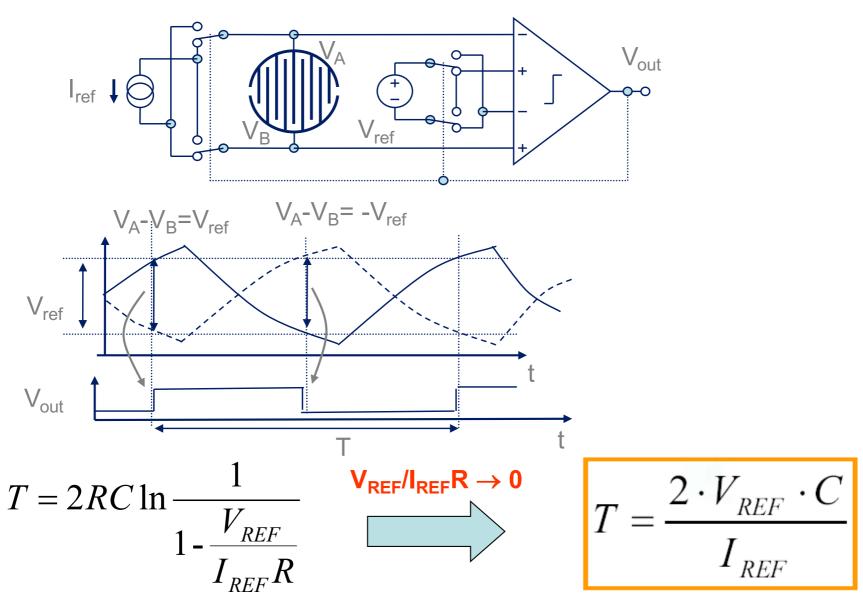
$$T = 2RC \ln \left[ 1 - \frac{V_{ref}}{RI_{ref}} \right]^{-1}$$

$$T = 2RC \ln \left[ \frac{1}{1 - \frac{V_{ref}}{RI_{ref}}} \right] \approx 2RC \ln \left[ 1 + \frac{V_{ref}}{RI_{ref}} \right] \approx \frac{2CV_{ref}}{I_{ref}}$$

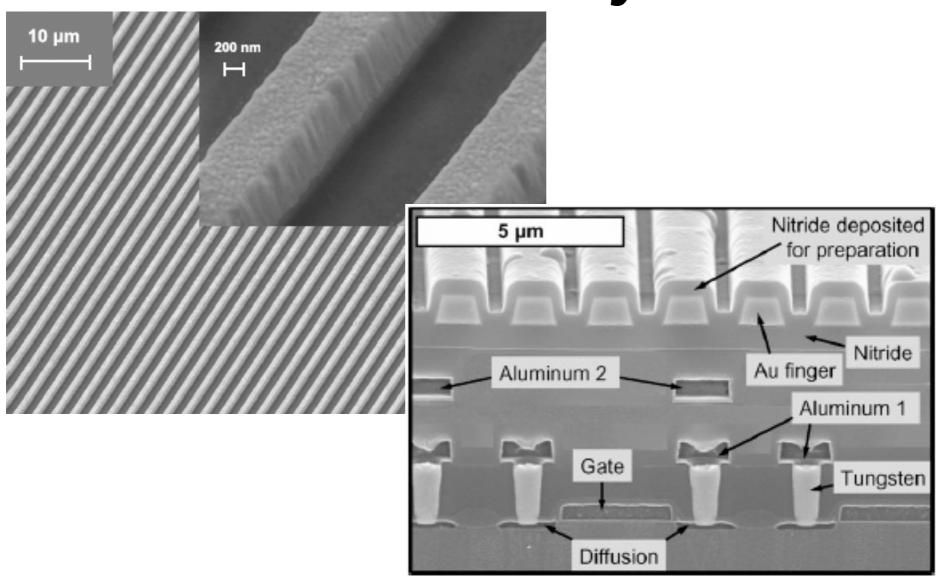
Method for the estimation of the Capacitance

### Frequency to Capacitance Measurement (FTCM)

Principle: Frequency To Capacitance Mode

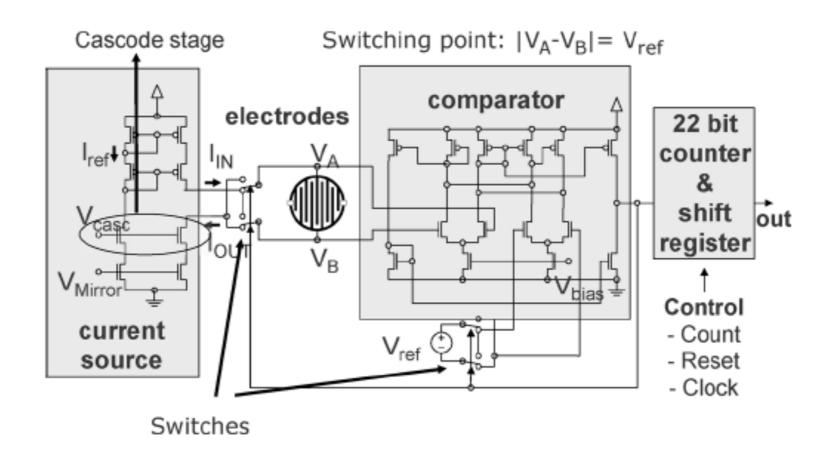


### Electrodes Layout



(c) S.Carrara

### **Chip Architecture (FTCM)**



### Measurements Set-up

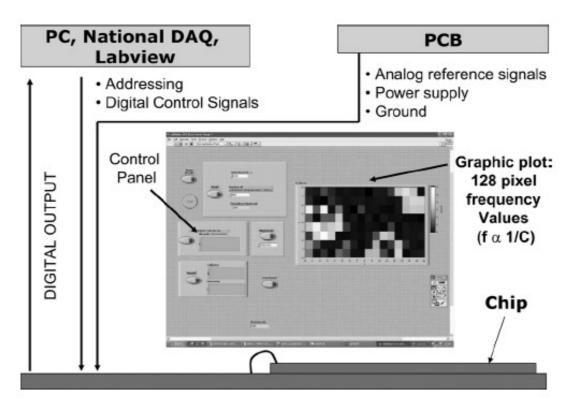


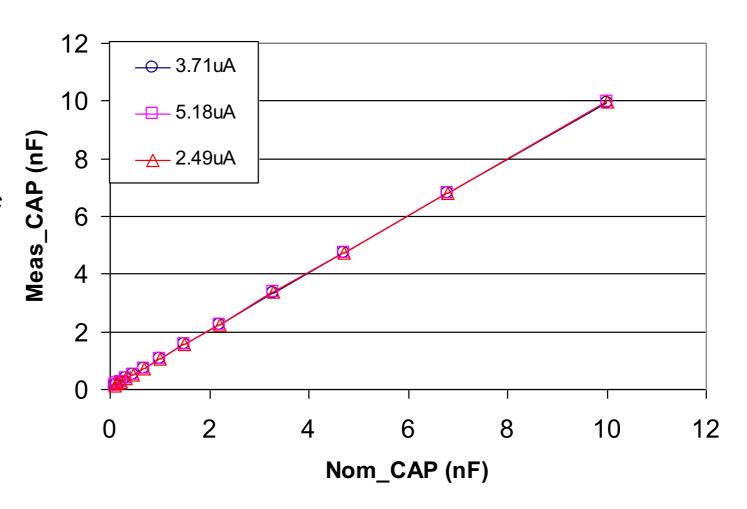
Fig. 9. Schematic representation of the measurement set-up. Voltage reference signals and power supply are generated by circuitry on the PCB. Digital control signals are provided by a PC. The LabView interface manages all the parameters involved in the measurements and shows directly on the screen the measurement results of the whole array.

#### Validation Test

A test structure has been implemented on chip beside the array to characterize the measurement circuit with discrete test capacitances (10 pF -10 nF)

Slope = 0.9837Intercept = 62 pF $\sigma < 0.3 \%$ 

Offset is due to parasitic capacitances of cables



### Probes property on FTCM mode

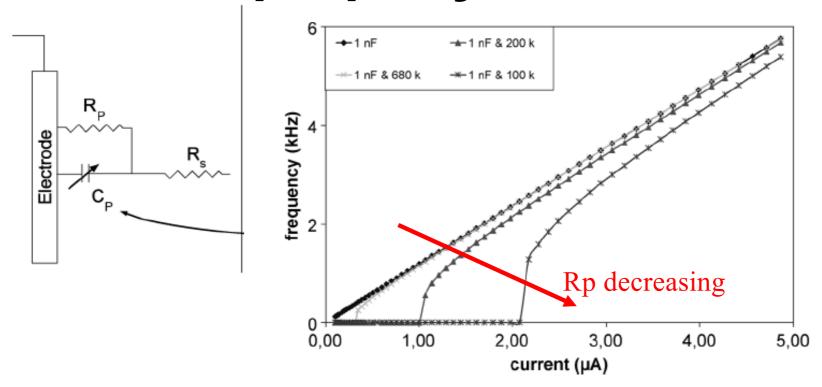
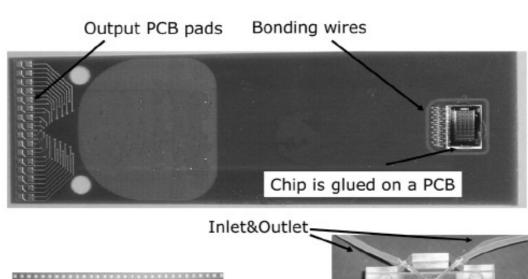
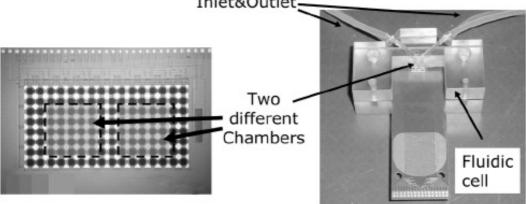


Fig. 12. Frequency versus reference current showing that a significant influence of the parallel resistance on the measurement result occurs only at low current values and at  $R_P$  values lower than 680 k $\Omega$ .

The linearity between the current and the measured frequency is lost at low current if the CMOS/Bio interface is not a perfect capacitor

### Liquid Measurement set-up





### **DNA** detection in FTCM mode

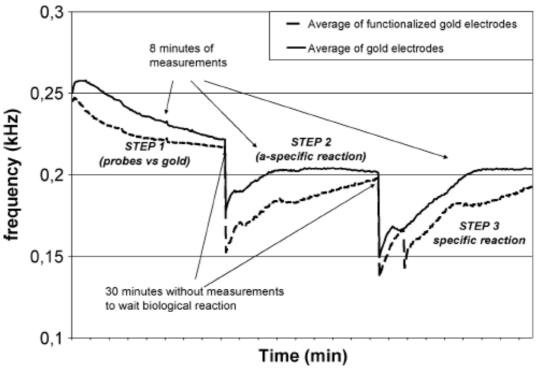


Fig. 13. Frequency changes of the average of reference electrodes (continuous line), and the average of functionalized electrodes (dashed line) show a larger gap after DNA hybridization step considering the stable value reached at the end of the transient.

Time stability on the single chip-spot is pour due to nano-scale aperture in the probes surfaces

### **DNA** detection in FTCM mode

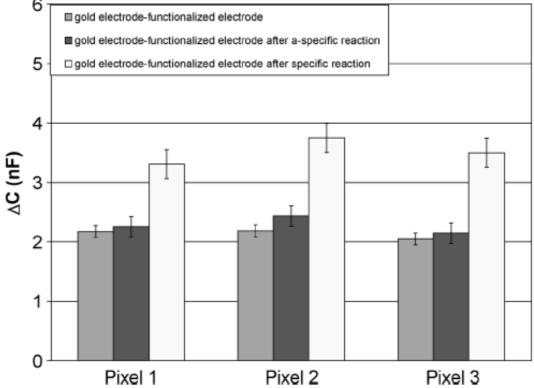
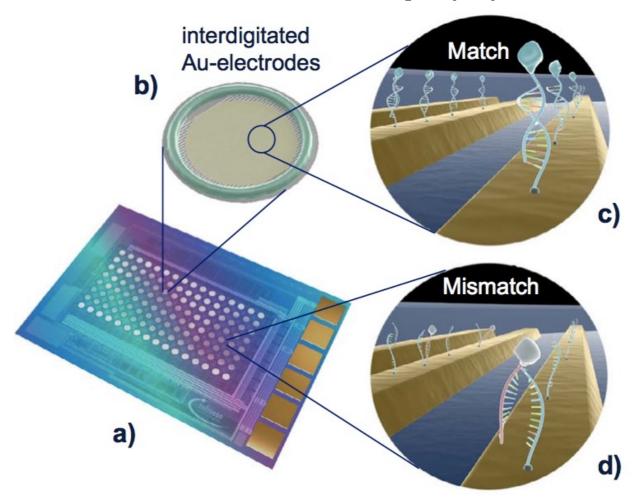


Fig. 14. Typical variations for several pixels among functionalized electrodes and the average value of reference gold electrodes. Capability to distinguish between specific and nonspecific binding is shown for each pixel.

In chip spot-by-spot reproducibility is improved due to better cleaning of the spot gold electrodes

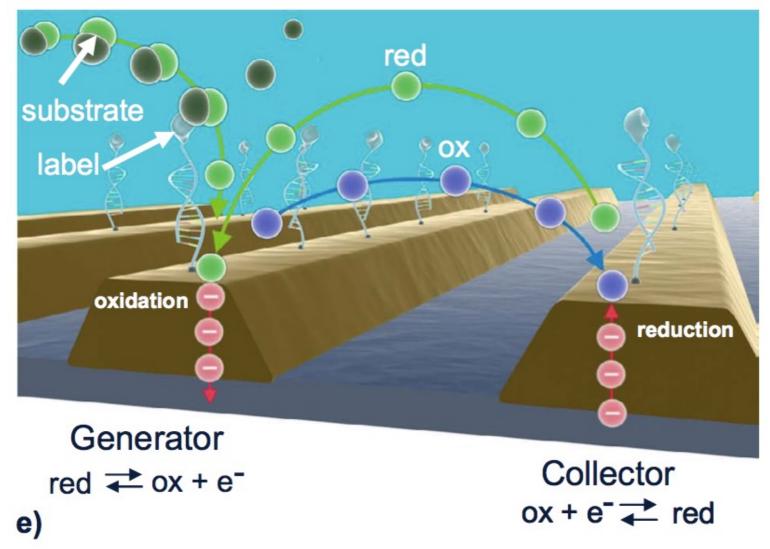
### **Amperometric Detection of DNA**

Figure by Frey et al, IEEE ISCAS 2015



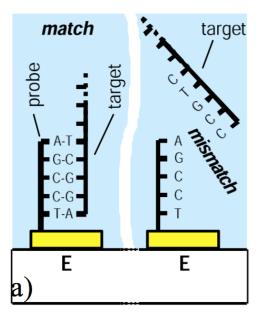
Electrochemical labels might be used to detect DNA

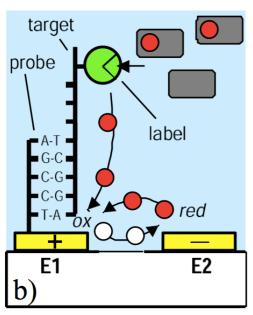
### **Amperometric Detection of DNA**



Redox species can be then measured at the electrodes

### **Amperometric Detection Principle**



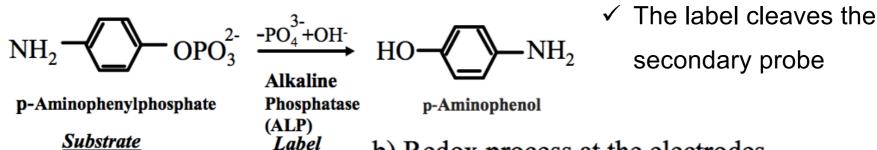


#### **How it works:**

- ✓ First, single stranded DNA molecules (about 20 bases) are immobilized by using a spotting machine on top of the gold electrodes due to gold-thiol coupling.
- ✓ Then, the chip is flooded with an analyte containing
  labeled target DNA ss: hybridization takes place in case
  of matching.
- ✓ A suitable substrate is applied to the buffer solution and
  it is enzymatically cleaved by the label.
- ✓ Resulting species starts an electrochemical redox process at the electrodes.
- ✓ Faradaic currents generated by the related redox process is detected and transduces DNA hybridization

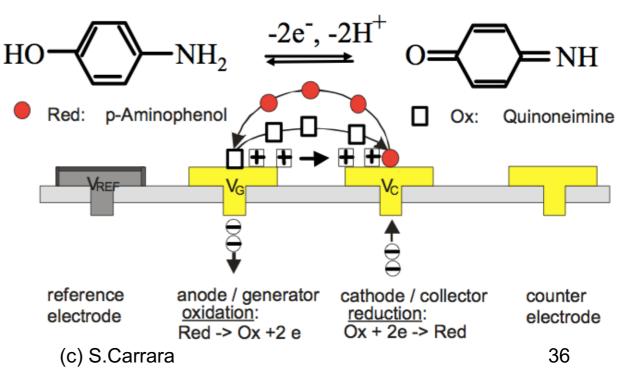
### Enzymatically cleavage & redox process

a) Process at the label

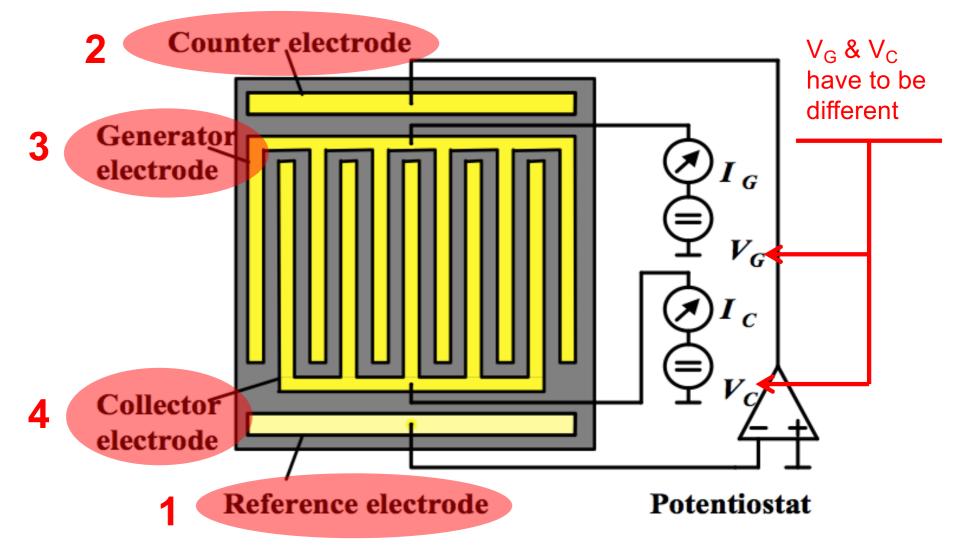


✓ The product of the cleavage is generating an oxidation process at the anode and, once oxidized, a reduction at the cathode

b) Redox process at the electrodes

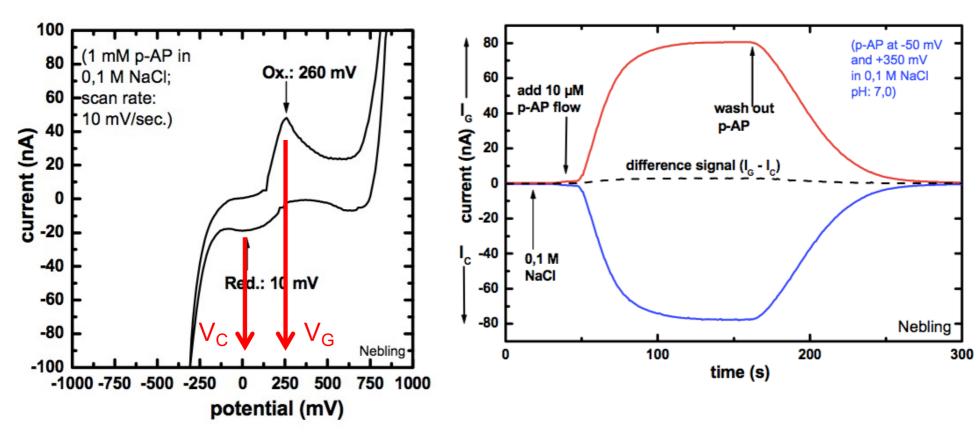


### The Electrochemical Cell



A 4-electrode Electrochemical Cell is here required

# **Cyclic Voltammetry**

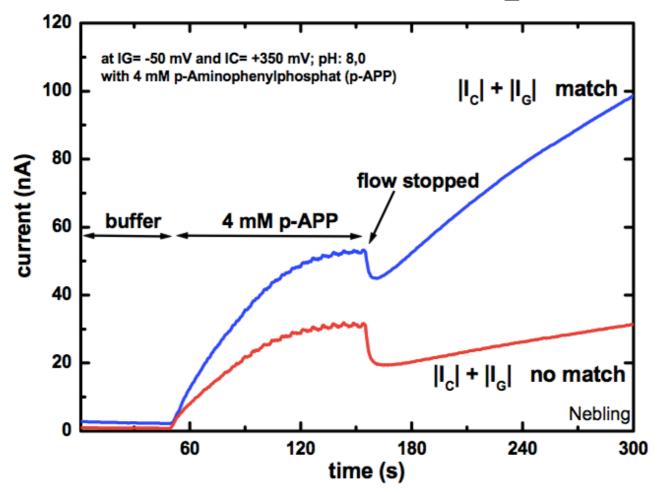


✓ Typical Cyclic Voltammetry acquired with 3-electrode cell

✓ Chronoamperometry acquired with 4-electrode cell

Simultaneous acquisition of Ox/Red current with 4-el

### Match/Mismatch DNA Hybridization

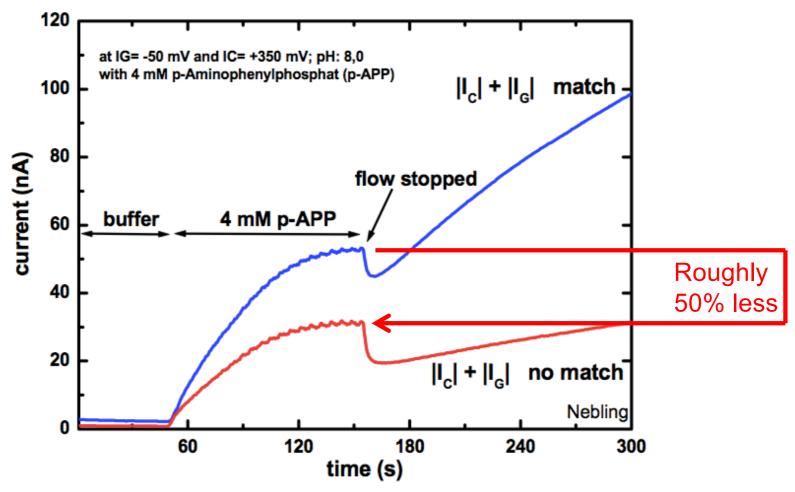


Successful detection of the matching sequence by significant signal by non-matching ones too

Gibbs free energy for Match/Mismatch

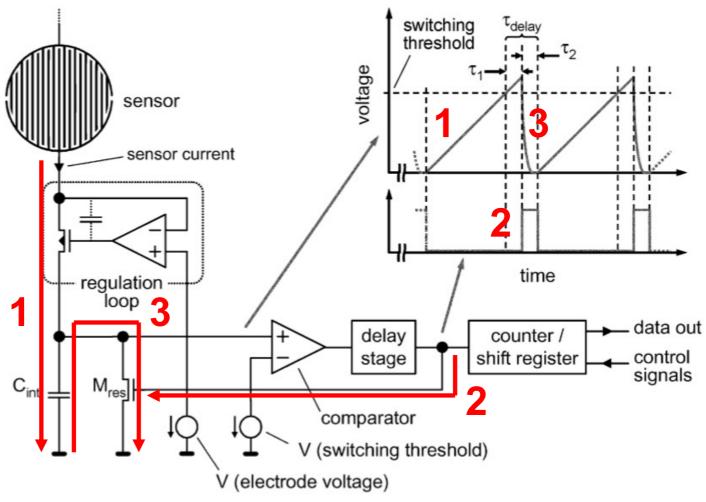
duplex	Experimental  △G [kJ/mol]	
GGTTATTGG	-26.8 ——	
CCAATAACC		
GGTTCTTGG	-31.4	
CCAAGAACC		Roughly
GGTTTTTGG	-29.5	50% less
CCAAAAACC		
GGTTATTGG	-12.0	
CCAAAAACC		
GGTT <mark>C</mark> TTGG	-12.4	
CCAATAACC		
GGTTTTTGG	S.Carrara	
CCAAGAACC (c)	S. Callala	

## Match/Mismatch DNA Hybridization



Successful detection of the matching sequences but significant signal by non-matching too

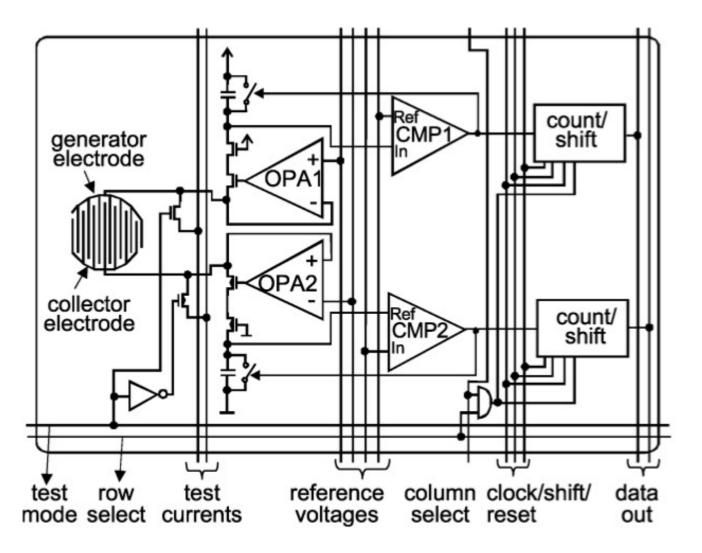
### **Current CMOS Readout**



Frequency-To-Current Conversion (FTCC) method is used here too

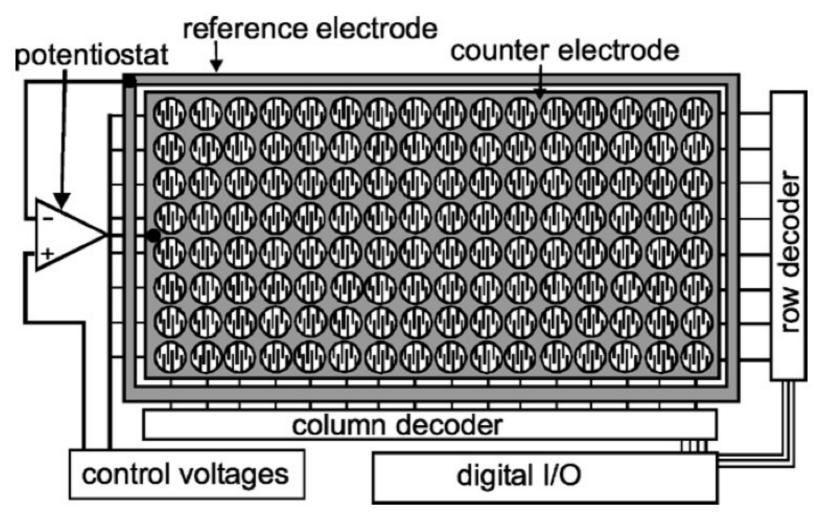
(c) S.Carrara

### **Current CMOS Readout**



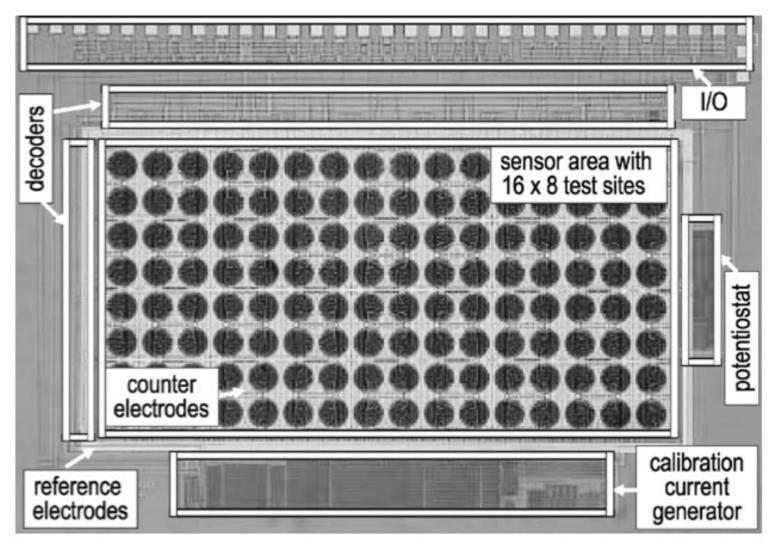
Sensor-site circuit architecture with digital output

# **Array Architecture**



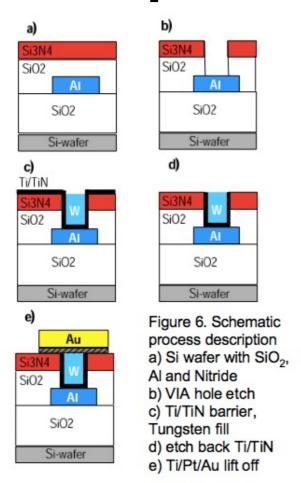
Whole Chip architecture including Row/Column decoders

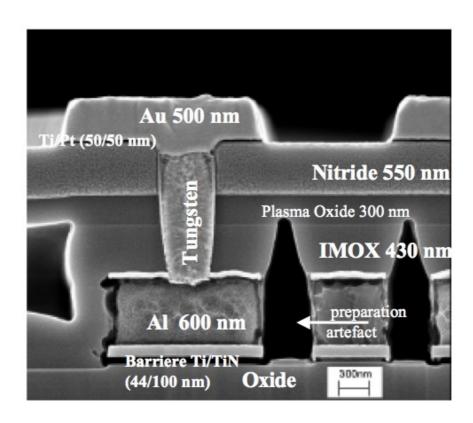
### The realized IC



Chip microphotograph. Total dimensions are 6.4 x 4.5 mm<sup>2</sup>.

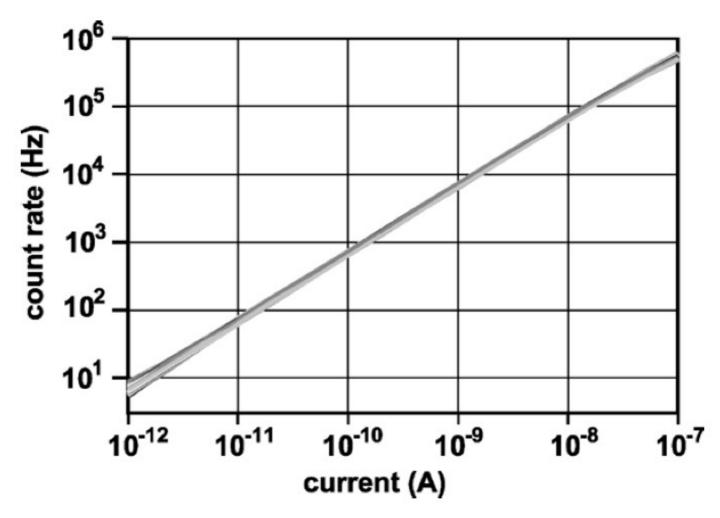
### **Exposed IC-Die Electrodes**





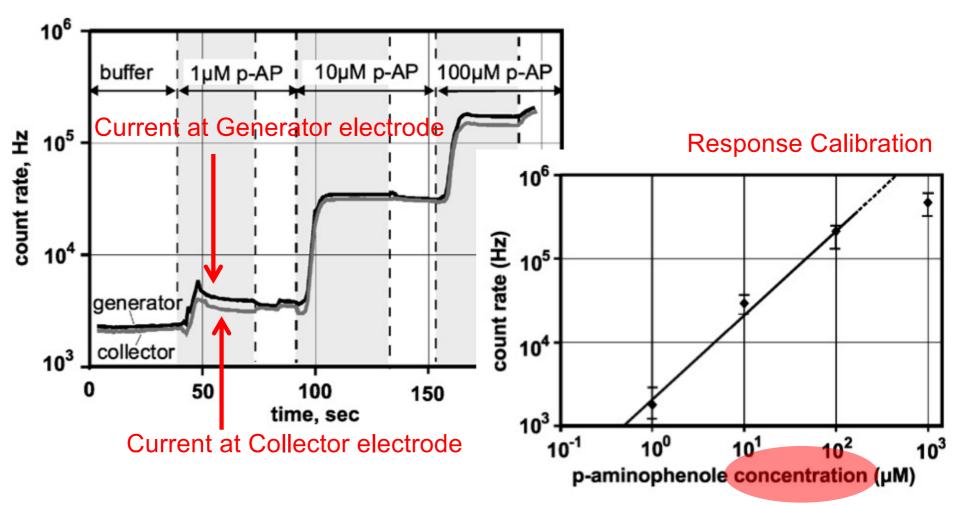
Electrochemical electrodes are created on top of the last CMOS metal Al layer

## **Frequency Readout**



Measured count rate of all 128 DNA sensors

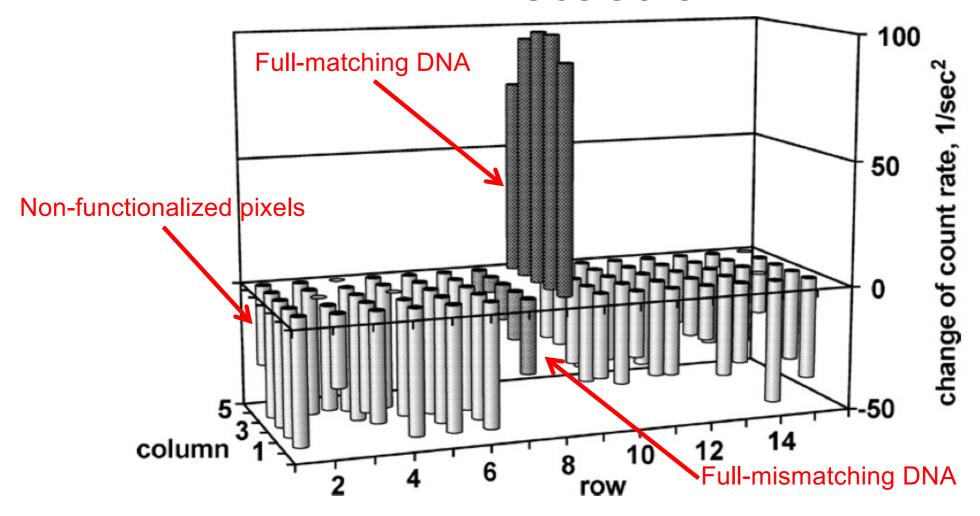
### **Test Measures**



Concentration of the secondary probe, not of the DNA

Response of the sensor versus secondary probe

### **DNA Detection**



Row 8: full matching DNA, row 7: full mismatching DNA, all other positions not functionalized.